# Towards a Compact Fiber Laser for Multimodal Imaging

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**Abstract:** We report on multimodal depth-resolved imaging of unstained living Drosophila Melanogaster larva using sub-50 fs pulses centered at 1060 nm wavelength. Both second harmonic and third harmonic generation imaging modalities are demonstrated.
**OCIS codes:** (170.5810) Scanning microscopy; (180.4315) Nonlinear microscopy; (180.6900) Three-dimensional microscopy.

## 1. Introduction

Due to the benefits of high contrast ratio, sub-micrometer resolution and depth resolved imaging multiphoton microscopy has been proven to be a powerful tool for studying living tissues [1, 2]. Especially for second harmonic generation (SHG) or third harmonic generation (THG) microscopy, no sample labeling is needed, which makes those methods preferable for non-invasive in vivo tissue imaging. In addition, SHG and THG provide complementary information due to their different optical-response mechanism. For both SHG and THG imaging, ultrashort laser pulses are preferred to achieve good multiphoton efficiency. It is found that SHG or THG efficiency is inversely proportional to the pulse duration or pulse duration square, respectively [3-6]. For clinical use, a compact and environmentally stable laser is need. In the past decade, fiber lasers have emerged as ideal ultrafast light sources [7]. Here an Yb fiber oscillator [8], capable of delivering pulses as short as ~50 fs at 1060 nm central wavelength, is tested for multiphoton microscopy imaging. The capability of this laser for multiphoton microscopy is evaluated with different samples including prepared slides with stained mouse kidney and mouse intestine sections and unstained living whole Drosophila Melanogaster larva. Images generated by different modalities such as two-photon excited fluorescence (TPEF), SHG and THG are compared. Depth scan of SHG and THG is conducted and reconstructed 3D images are shown.

## 2. Experimental Setup

An Yb fiber oscillator is operated at 43 MHz with average power up to 400 mW. This laser is based on an all-normal dispersion cavity and is similar to the design of the laser described in ref. [8]. The output laser beam is guided through a 4-f folded pulse shaper (MIIPS Box 640, Biophotonic Solutions), which is used to compensate the high-order (second order and higher) phase distortions to deliver transform limited pulses at the focal plane. Output from the pulse shaper is directed to a laser-scanning multiphoton microscope. The laser beam is scanned by a galvanometer mirrors (QuantumDrive-1500, Nutfield Technology, Inc.) and coupled into a water-immersed objective (Zeiss LD C-APOCHROMAT 40x/1.1). The generated SHG and TPEF emissions from samples are collected in the Epi direction, being filtered out using a dichroic mirror (700DCSPXR, Chroma Technology Corp.) and a short-pass emission filter (ET680-SP-2P8, Chroma Technology Corp.). A photomultiplier (PMT, HC20-05MOD, Hamamatsu) is used to collect the SHG/TPEF signal. THG, which is primarily generated in the forward direction, is collected by a UV compatible objective (HP ReflX, NT59-886, NA 0.28, Edmund Optics). The THG signal is also separated from the excitation light by a 400 nm short pass filter and detected by a PMT (H10720-210, Hamamatsu) whose signal is amplified (SRS445, Stanford Research Systems). The focal plane is moved to different layers using a step motor capable of making precisely controlled 2 μm height steps. All the SHG or THG images are then incorporated into a 3-D image.

## 3. Experimental Results

Excitation laser pulses are compressed to about 50 fs at the focal plane of objective using the MIIPS enabled pulse shaper. After pulse compression, samples are loaded onto the focal plane of the objective. To calibrate the microscope, two stained commercial samples (mouse kidney and mouse intestine, Molecular Probes) that have uniform thickness are imaged. For these two samples, the signal detected in Epi direction is mainly from two- or three-photon excited fluorescence. On the forward direction, mainly THG/three-photon excited fluorescence signal is detected. By combining the signal from Epi and forward directions, it is clearly seen that they provide complementary information for each other (see Fig. 1).

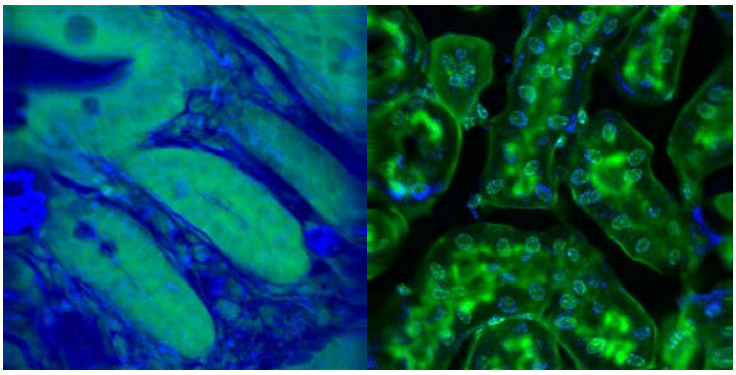

Fig. 1. Composition of TPEF (green, false color) and THG (blue, false color) imaging of mouse intestine (left) and mouse kidney (right), 150 μm x150 μm area represented.

Beyond imaging pre-labeled samples, depth-resolved imaging of unstained live tissue is of greater importance. In a previous report [3], we demonstrated a fiber laser delivering 30 fs pulses used for multiphoton imaging of living tissues. However, the low pulse energy (about 1 nJ) limited the imaging depth capability. The laser used in this work provides 10 times greater pulse energy and only slightly longer pulse duration. Depth resolved images of third instar Drosophila larva are shown in Fig.2. The THG 3D image shows many more structures, for example the adipose tissue in the lower left corner, and less scattering than the SHG. The total scanned depth is about 90 μm. The videos of the reconstructed SHG and THG 3D images will be demonstrated.

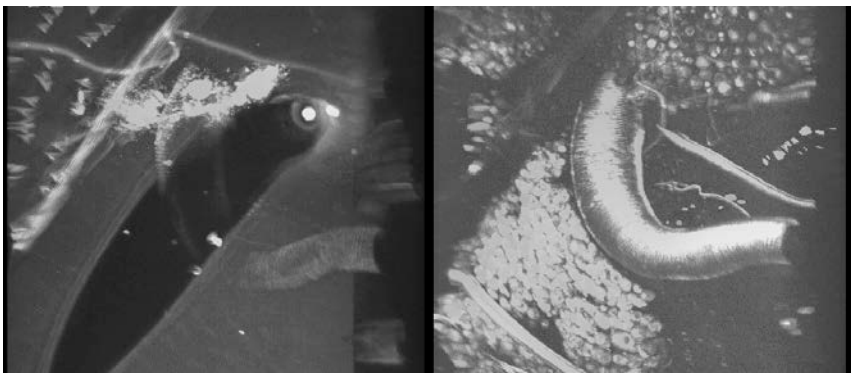

Fig. 2. Projection of 3-D images at 30° angle for SHG (left) and THG (right) microscopy of the third instar D. Melanogaster larva. Images are of the same 150 μm x150 μm region centered at the trachea, but different contrast mechanisms highlight different organs.

## 4. Conclusion

In conclusion, we have demonstrated live depth resolved imaging of Drosophila larva using a 50 fs Yb fiber laser. The shorter pulse durations achieved by the laser greatly enhance two- and three-photon induced modalities in both stained and unstained living tissues.

## 5. References

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